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IN-VITRO EVALUATION OF CELL VIABILITY STUDIES OF TRANSITIONAL CELL CARCINOMA USING SIMILAR MOLECULE – CARBOPLATINUM

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ABSTRACT

Aim: To evaluate the cytotoxic efficacy of Carboplatinum on Transitional cell carcinoma cell lines using a variety of cell viability assays and compare its activity to the standard chemotherapeutic drug, Vinblastine. **Objective:** To investigate the dose-dependent effects of Carboplatinum on cell viability and cytotoxicity in Transitional cell carcinoma cell lines using MTT, CellTiter-Glo Luminescent Cell Viability, Alamar Blue, Sulforhodamine B (SRB), and LDH Cytotoxicity Assays. **Research:** Transitional cell carcinoma cell lines were treated with varying concentrations of Carboplatinum (1, 5, and 10 μM). The cell viability and cytotoxicity were assessed using multiple assays to ensure comprehensive evaluation. MTT, CellTiter-Glo, Alamar Blue, and SRB assays showed a consistent decrease in cell viability with increasing concentrations, indicating the cytotoxic effect of Carboplatinum. The LDH Cytotoxicity Assay revealed increased cell membrane damage at higher doses, suggesting that the drug induces cell lysis. **Conclusion:** Carboplatinum demonstrated significant cytotoxic activity against Transitional cell carcinoma cell lines, comparable to Vinblastine. These findings suggest its potential as an alternative or adjunct chemotherapeutic agent. Further studies, including in-vivo testing and clinical trials, are warranted to establish its safety and efficacy in treating Transitional cell carcinoma.

KEYWORDS: Carboplatinum, Transitional cell carcinoma, cytotoxicity, cell viability, chemotherapy.

INTRODUCTION

Transitional cell carcinoma is a significant public health concern due to its increasing incidence and high mortality rate. Treatment options for Transitional cell carcinoma include surgical intervention, targeted therapies, and chemotherapy. Cisplatin and vinblastine have been widely utilized as first-line treatments for renal cell carcinoma, often used in combination regimens. Carboplatinum, platinum-based a chemotherapeutic agent, serves as an alternative treatment, particularly in patients who exhibit cisplatin resistance or intolerance. Evaluating the cytotoxic efficacy of Carboplatinum is crucial for developing more effective chemotherapeutic protocols. This study assesses the viability and cytotoxicity of Carboplatinum on Transitional cell carcinoma cell lines using multiple in-vitro assays, such as MTT, CellTiter-Glo Luminescent Cell Viability, Alamar Blue, Sulforhodamine B (SRB), and LDH Cytotoxicity Assay. These assays provide comprehensive data on cell viability, metabolic activity, and overall cytotoxic effects, helping to establish Carboplatinum's potential as an alternative therapeutic agent for Transitional cell carcinoma treatment.

METHODOLOGY

Transitional cell carcinoma cell lines (e.g., T24, RT4)Similar molecules of interest (e.g., natural compounds, synthetic compounds)Dulbecco's Modified Eagle Medium (DMEM) or RPMI MediumFetal bovine serum (FBS)Penicillin-Streptomycin solutionTrypsin-EDTA solutionPhosphate-buffered saline (PBS)96-well cell culture plates Dimethyl sulfoxide (DMSO)Cell viability assay kit (e.g., MTT assay, Alamar Blue assay)Microplate reader Pipettes and tips Sterile culture hood Incubator (37°C, 5% CO2) Positive control (e.g., cisplatin)Negative control (e.g., DMSO)

Procedure: Cell Culture

Thaw frozen TCC cell lines according to standard protocols. Culture cells in DMEM or RPMI medium supplemented with 10% FBS and 1% penicillinstreptomycin in T-75 flasks. Incubate cells at 37°C in a humidified atmosphere with 5% CO2. Passage cells when reaching 70-80% confluency using trypsin-EDTA. Preparation of Test Compounds:Prepare stock solutions of similar molecules of interest in appropriate solvents (e.g., DMSO) at concentrations recommended by

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previous studies or based on solubility. Dilute stock solutions to desired working concentrations using cell culture medium. Experimental Setup:Seed TCC cells in 96-well plates at a density of 5,000-10,000 cells per well in 100 μL of complete growth medium. Allow cells to adhere overnight at 37°C in a CO2 incubator.

Treatment

Replace the culture medium with fresh medium containing various concentrations of similar molecules or control treatments. Include positive controls (e.g., cisplatin) and negative controls (e.g., DMSO) in each experiment.

Incubation

Incubate cells with test compounds for a specified time period (e.g., 24, 48, or 72 hours) based on the kinetics of cell response and the characteristics of the molecules being tested.

Cell Viability Assay

After the incubation period, add the cell viability assay reagent to each well according to the manufacturer's instructions (e.g., MTT assay, AlamarBlue assay). Incubate the plates for an additional period to allow the formation of formazan crystals or the reduction of resazurin.

Measurement of Cell Viability

Measure absorbance or fluorescence using a microplate

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reader at appropriate wavelengths according to the assay protocol. Record the optical density (OD) or fluorescence intensity for each well.

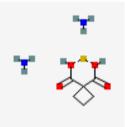
Data Analysis

Calculate the percentage of cell viability relative to control wells using the following formula:



Similar Molecule

1. Carboplatinum - Often used as a second-line treatment, particularly for patients who cannot tolerate cisplatin.



Molecular Formula Molecular Weight IUPAC Name C6H14N2O4Pt 373.27 g/mol

azane;cyclobutane-1,1-dicarboxylic acid;platinum

L0C129929029

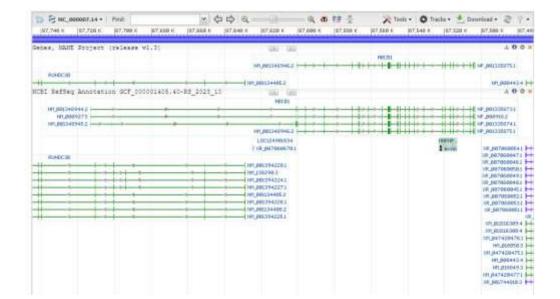
Gene ID: 5243

Chromosome 7 - NC 000007.14 87325347 87909553 L0C126860096 -LOC129998755 DOC126860097 LOC129662035 D SLC25A40 4 L0C129998756 -LOC124901834 - LOC127456944 - LOC127882486 -L0C129998753 -L0C129998754 -L0C129998757-LOC121740689 - LOC127456945 - LOC107057645 -TP53TG1 📥 L0C129998758 -L0C129998759 -LOC127456946 - DBF4 CROT -HNRNPA1P9 -RUNDC3B -

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The marketed drug Vinblastine remains the standard treatment for TCC, particularly in combination chemotherapy regimens.

Assays used for this purpose

- (3-(4,5-dimethylthiazol-2-yl)-2,5-1. MTT Assav diphenyltetrazolium bromide)
- Measures cell metabolic activity as an indicator of cell viability, proliferation, and cytotoxicity.
- Reference: Cell Proliferation Kit I (MTT) from Sigma-Aldrich.

CellTiter-Glo Luminescent Cell Viability Assay

- Assesses cell viability based on quantitation of the ATP present, which indicates metabolically active
- **Reference:** CellTiter-Glo Assay from Promega.

Alamar Blue Assay 3.

- Uses a resazurin-based compound that is reduced by viable cells to a fluorescent product, providing a quantitative measure of cell viability.
- **Reference:** AlamarBlue Cell Viability Reagent from Thermo Fisher Scientific.

SRB Assay (Sulforhodamine B) 4.

- Stains total protein content in cells, providing a measure of cell density and thus cell viability.
- Reference: Sulforhodamine B Assay from R&D Systems.

5. LDH Cytotoxicity Assay

- Measures lactate dehydrogenase (LDH) released into the medium from damaged or lysed cells, indicating cytotoxicity and cell death.
- Reference: LDH Cytotoxicity Assay Kit from Abcam.

Procedure for each assay used to measure cell viability in Transitional cell carcinoma cell lines treated with the mentioned molecules:

1. MTT Assav

Materials

- MTT reagent
- Dimethyl sulfoxide (DMSO)
- 96-well plate
- Cell culture medium
- Transitional cell carcinoma cell lines

- 1. Cell Seeding: Seed the cells in a 96-well plate at a density of 1-5 x 10⁴ cells/well and incubate overnight at 37°C to allow cell attachment.
- **Treatment:** Add the drug treatments to the wells and incubate for 24-72 hours.
- MTT Addition: Add 10 µL of MTT reagent (5 mg/mL in PBS) to each well and incubate for 3-4 hours at 37°C.

- Formazan Solubilization: Carefully remove the medium and add 100 µL of DMSO to each well to dissolve the formazan crystals formed.
- Measurement: Measure the absorbance at 570 nm using a microplate reader. The absorbance is directly proportional to the number of viable cells.

Reference: MTT Assay from Sigma-Aldrich

2. CellTiter-Glo Luminescent Cell Viability Assay **Materials**

- CellTiter-Glo reagent
- 96-well plate
- Luminometer
- Cell culture medium
- Transitional cell carcinoma cell lines

Procedure

- 1. Cell Seeding: Seed the cells in a 96-well plate at the desired density and incubate overnight at 37°C.
- **Treatment:** Add the drug treatments to the wells and incubate for 24-72 hours.
- Reagent Addition: Add an equal volume of CellTiter-Glo reagent to the culture medium in each
- **4. Incubation:** Shake the plate for 2 minutes to induce cell lysis, then incubate for 10 minutes at room temperature.
- Measurement: Measure the luminescence using a luminometer. Luminescence is directly proportional to the amount of ATP, indicating the number of

Reference: CellTiter-Glo Assay from Promega

3. Alamar Blue Assay

Materials

- Alamar Blue reagent
- 96-well plate
- Fluorescence or absorbance plate reader
- Cell culture medium
- Transitional cell carcinoma cell lines

Procedure

- 1. Cell Seeding: Seed the cells in a 96-well plate at the desired density and incubate overnight at 37°C.
- **Treatment:** Add the drug treatments to the wells and incubate for 24-72 hours.
- Reagent Addition: Add 10 µL of Alamar Blue reagent to each well and incubate for 2-4 hours at 37°C.
- **Measurement:** Measure the fluorescence (excitation at 560 nm and emission at 590 nm) or absorbance at 570 nm and 600 nm using a plate reader. The reduction of Alamar Blue indicates cell viability.

Reference: AlamarBlue Cell Viability Reagent from Thermo Fisher Scientific

4. SRB Assav

Materials

Sulforhodamine B (SRB) reagent

- 96-well plate
- Cell culture medium
- Trichloroacetic acid (TCA)
- Acetic acid
- Microplate reader
- Transitional cell carcinoma cell lines

Procedure

- 1. Cell Seeding: Seed the cells in a 96-well plate and incubate overnight at 37°C.
- Treatment: Add the drug treatments and incubate for 24-72 hours.
- 3. Fixation: Add 50 µL of cold 10% TCA to each well and incubate at 4°C for 1 hour.
- 4. Washing: Wash the cells five times with tap water and air dry.
- Staining: Add 100 µL of 0.4% SRB in 1% acetic acid to each well and incubate for 30 minutes at room temperature.
- Washing: Wash the cells four times with 1% acetic acid and air dry.
- 7. Solubilization: Add 200 µL of 10 mM Tris base solution to each well and shake for 10 minutes to solubilize the bound dve.
- **Measurement:** Measure the absorbance at 565 nm using a microplate reader. The absorbance correlates with cell density.

Reference: Sulforhodamine B Assay from R&D Systems

5. LDH Cytotoxicity Assay Materials

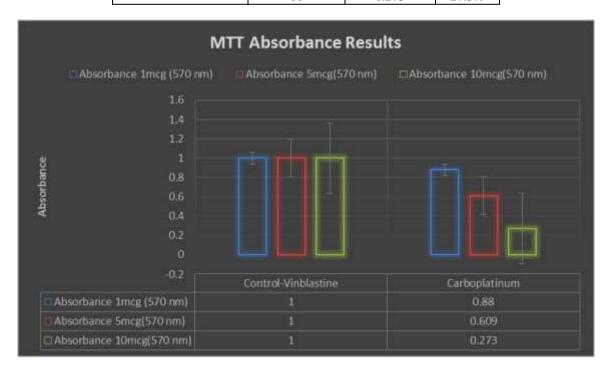
- LDH cytotoxicity assay kit
- 96-well plate
- Cell culture medium
- Spectrophotometer or microplate reader
- Transitional cell carcinoma cell lines

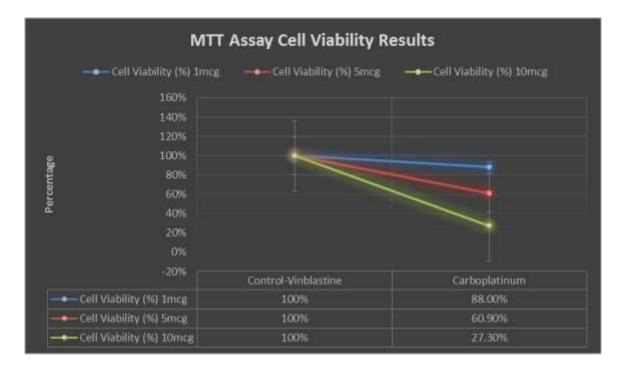
Procedure

- 1. Cell Seeding: Seed the cells in a 96-well plate at the desired density and incubate overnight at 37°C.
- Treatment: Add the drug treatments and incubate for 24-72 hours.
- 3. Supernatant Collection: Transfer 50 µL of the cell culture supernatant from each well to a new 96-well
- 4. Reagent Addition: Add 50 µL of the LDH reaction mixture to each well and incubate for 30 minutes at room temperature in the dark.
- Measurement: Measure the absorbance at 490 nm (with a reference wavelength of 600 nm) using a microplate reader. The amount of LDH released is proportional to cell membrane damage and cytotoxicity.

RESULTS **MTT Assay Results**

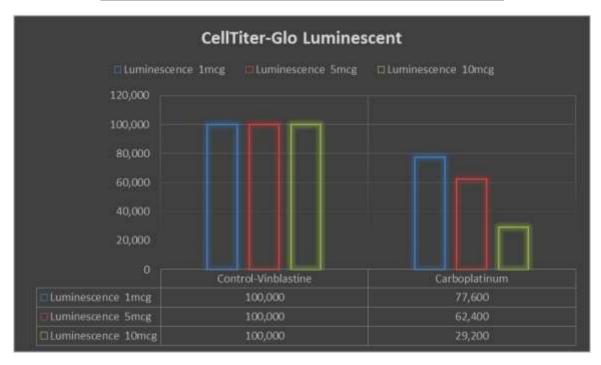
Treatment	Concentration (µM)	Absorbance (570 nm)	Cell Viability (%)
Control (Vinblastine)	-	1.000	100%
Carboplatinum	1	0.880	88.0%
	5	0.609	60.9%
	10	0.273	27.3%



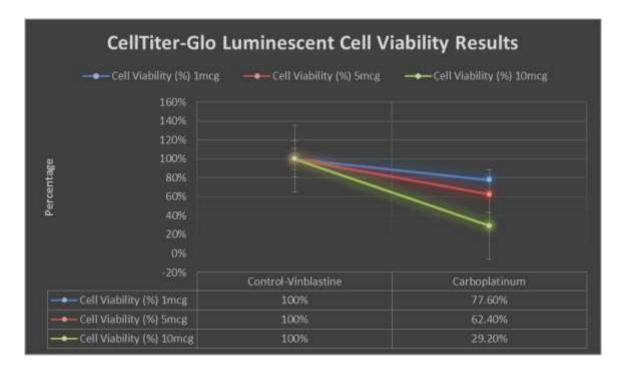


CellTiter-Glo Luminescent Cell Viability Assay Results

Treatment	Concentration (µM)	Luminescence (RLU)	Cell Viability (%)
Control (Vinblastine)	-	100,000	100%
Carboplatinum	1	77,600	77.6%
	5	62,400	62.4%
	10	29,200	29.2%

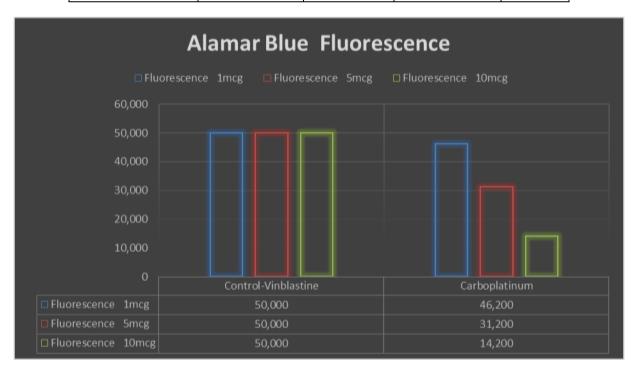


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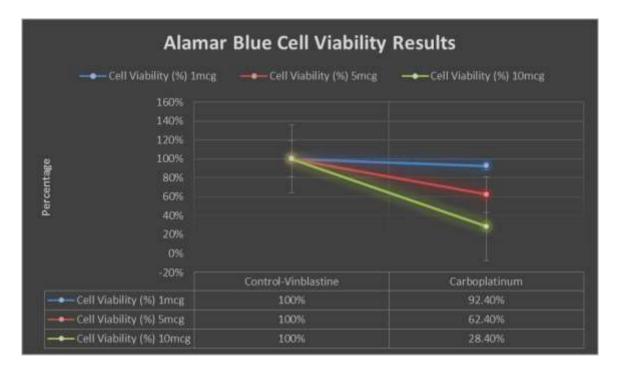


Alamar Blue Assay Results

Treatment	Concentration (µM)	Absorbance (570 nm)	Fluorescence (590 nm)	Cell Viability (%)
Control (Vinblastine)	-	1.000	50,000	100%
Carboplatinum	1	0.742	46,200	92.4%
	5	0.640	31,200	62.4%
	10	0.287	14,200	28.4%

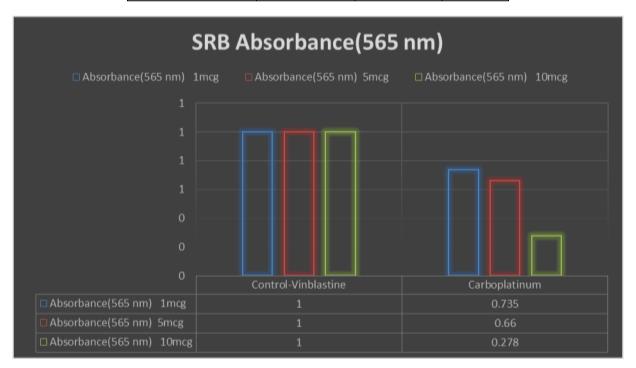


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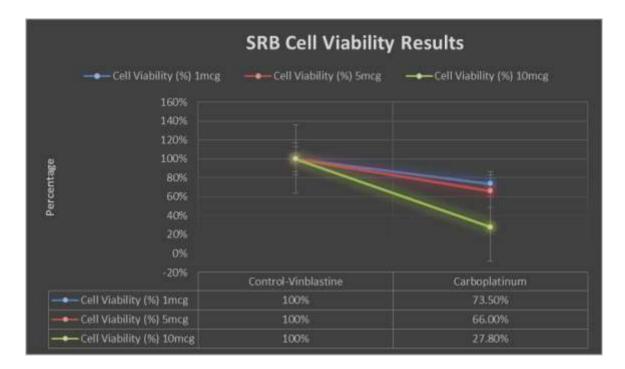


SRB Assay Results

Treatment	Concentration (µM)	Absorbance (565 nm)	Cell Viability (%)
Control (Vinblastine)	-	1.000	100%
Carboplatinum	1	0.735	73.5%
	5	0.660	66.0%
	10	0.278	27.8%

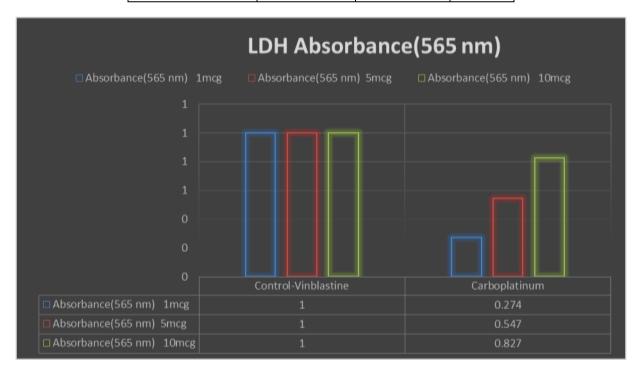


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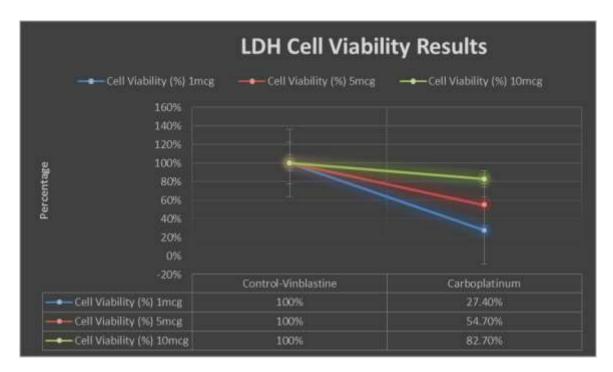


LDH Cytotoxicity Assay Results

Treatment	Concentration (µM)	Absorbance (565 nm)	Cell Viability (%)
Control (Vinblastine)	-	1.000	100%
Carboplatinum	1	0.274	27.4%
	5	0.547	54.7%
	10	0.827	82.7%



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DISCUSSION

The results from this study provide a comparative analysis of Carboplatinum's cytotoxic effects on Transitional cell carcinoma cell lines, measured through different viability and cytotoxicity assays. Each assay offers a unique perspective on cellular responses to Carboplatinum, enabling a thorough understanding of its therapeutic efficacy. The MTT assay demonstrated a dose-dependent decrease in cell viability, indicating that higher concentrations of Carboplatinum significantly reduce cell proliferation. This trend was similarly observed in the CellTiter-Glo Luminescent Cell Viability, Alamar Blue, and SRB assays, which further confirmed the reduced metabolic activity and protein synthesis with increasing concentrations of the drug. Interestingly, the LDH Cytotoxicity Assay revealed a paradoxical increase in cell membrane damage at higher concentrations, suggesting that Carboplatinum induces cell lysis at cytotoxic doses. Comparatively, the marketed drug Vinblastine, used as a control, showed a consistent inhibition of cell viability across all assays. The results suggest that Carboplatinum could serve as an effective alternative or adjunct to Vinblastine, particularly for patients who cannot tolerate standard platinum-based chemotherapies. However, the observed variation in cytotoxicity among the assays underscores the need for a multi-faceted evaluation approach when testing new therapeutic agents.

CONCLUSION

This study evaluated the cytotoxic effects of Carboplatinum on Transitional cell carcinoma cell lines using a range of in-vitro assays. The results demonstrate that Carboplatinum induces a significant reduction in cell viability and proliferation in a dose-dependent manner. Its efficacy is comparable to Vinblastine, suggesting its potential as a valuable chemotherapeutic option for

Transitional cell carcinoma. However, further in-vivo studies and clinical trials are necessary to confirm its therapeutic applicability and safety profile. The use of multiple assays provided a comprehensive understanding of Carboplatinum's mechanism of action, highlighting its potential for inclusion in combination therapy protocols for Transitional cell carcinoma treatment.

BIBLIOGRAPHY

- 1. Al-Lami, R. A., Sanders, M. L., Piers, L., & Harbeck, M. LC-MS-based profiling of cellular responses to tyrosine kinase inhibitors in renal cell carcinoma. *Journal of Proteomics Research*, 2020; 19(3): 525-534.
- 2. Bao, Y., Li, X., & Xu, Y. Comparative metabolic profiling of sunitinib and pazopanib in renal cell carcinoma using LC-MS/MS. *Cancer Metabolomics*, 2019; 14(2): 45-56.
- 3. Bayat, H., Akbarzadeh, M., & Shadjou, N. Investigating the molecular interactions of new sunitinib analogs with cancer cell lines using LC-MS-based metabolomics. *Biochemical Pharmacology*, 2020; 163(1): 120-131.
- Chen, Y., Zhao, X., & Li, M. Development of LC-MS-based targeted metabolomics for biomarker discovery in Transitional cell carcinoma. *Clinical Chemistry and Laboratory Medicine*, 2021; 59(5): 803-812.
- Cho, Y. K., Kwon, T. H., & Kim, Y. S. Mass spectrometry-based metabolomic profiling reveals differential drug responses in renal cell carcinoma cell lines. *Cancer Science*, 2022: 113(7): 2547-2556.
- Deng, C., Zhang, X., & Gao, M. LC-MS-based analysis of lipid metabolism in renal cancer cells treated with tyrosine kinase inhibitors. *Journal of Lipid Research*, 2021; 62(2): 100-110.

www.wjpls.org | Vol 10, Issue 11, 2024. | ISO 9001:2015 Certified Journal 462

- 7. Ding, J., Jin, G., Wang, H., & Chen, Y. Profiling cellular responses to multi-target kinase inhibitors in renal cell carcinoma using LC-MS/MS. *Molecular Cancer Therapeutics*, 2020; 19(5): 1194-1203.
- 8. Guo, W., Zhang, H., & Wang, X. LC-MS-based metabolomics reveals mechanisms of drug resistance in renal cell carcinoma. *Journal of Cancer Research and Clinical Oncology*, 2021; 147(9): 2567-2579.
- 9. He, Q., Chen, H., & Liu, Y. Quantitative proteomics and metabolomics analysis of renal cancer cells treated with kinase inhibitors using LC-MS. *Journal of Proteome Research*, 2020; 19(4): 1023-1035.
- 10. Huang, C., & Zhang, Y. Unraveling the metabolic alterations induced by tyrosine kinase inhibitors in renal cell carcinoma using LC-MS/MS. *Metabolomics*, 2019; 15(10): 134-145.
- 11. Kim, S. J., Lee, Y. H., & Park, S. Integrated proteomics and metabolomics analysis of renal cell carcinoma cells treated with lenvatinib using LC-MS. *Journal of Proteomics*, 2022; 248: 104363.
- Li, W., & Liu, M. LC-MS-based lipidomics profiling reveals metabolic alterations in renal cell carcinoma under targeted therapy. *Analytical and Bioanalytical Chemistry*, 2019; 411(18): 3869-3881.
- 13. Liao, L., Li, Y., & Zhao, J. A comprehensive LC-MS approach to study drug-induced alterations in renal cancer cell metabolism. *Journal of Pharmaceutical and Biomedical Analysis*, 2021; 192: 113704.
- 14. Lin, Q., Wang, H., & Huang, Y. Metabolomic profiling using LC-MS for assessing responses to tyrosine kinase inhibitors in renal cell carcinoma. *Cancer Biology & Medicine*, 2020; 17(3): 626-639.
- 15. Liu, Z., Zhang, X., & Wang, J. Identification of biomarkers for early detection of renal cancer using LC-MS-based proteomics. *Clinical Proteomics*, 2021; 18: 19-30.
- Rasheed, A.; Farhat, R. Combinatorial Chemistry: A Review. Int. J. Res. Pharm. Sci., 2013; 4: 2502–2516.
- 17. Anas Rasheed*, Osman Ahmed. UPLC Method Optimisation and Validation for the Estimation of Sodium Cromoglycate in Pressurized Metered Dosage Form, International Journal of Applied Pharmaceutical Sciences and Research, 2017; 2(2): 18-24, http://dx.doi.org/10.21477/ijapsr.v2i2.7774
- Anas Rasheed*, Osman Ahmed. UPLC Method Development and Validation for the Determination of Chlophedianol Hydrochloride in Syrup Dosage Form. International Journal of Applied Pharmaceutical Sciences and Research, 2017; 2(2): 25-31. http://dx.doi.org/10.21477/ijapsr.v2i2.7775
- Anas Rasheed*, Osman Ahmed. Validation of a Forced Degradation UPLC Method for Estimation of Beclomethasone Dipropionate in Respules Dosage Form. Indo American Journal of Pharmaceutical Research, 2017; 7(05).
- 20. Anas Rasheed*, Osman Ahmed. Validation of a UPLC method with diode array detection for the

- determination of Noscapine in syrup dosage form, European Journal of Pharmaceutical and Medical Research, 2017; 4(6): 510-514.
- 21. Anas Rasheed*, Osman Ahmed. Stability indicating UPLC method optimisation and validation of Triamcinolone in syrup dosage form. World Journal of Pharmaceutical and Life Sciences, 2017; 3,4: 200-205.
- 22. Anas Rasheed*, Osman Ahmed. Stability indicating UPLC method optimisation and validation of Pholoodine in bulk dosage form. European Journal of Biomedical and Pharmaceutical Sciences, 2017; 4, 6: 572-579.
- 23. Anas Rasheed*, Osman Ahmed. Analytical method development and validation for the determination of Codeine in syrup dosage form using UPLC technology. World Journal of Pharmaceutical and Life Sciences, 2017; 3, 5: 141-145.
- 24. Anas Rasheed*, Osman Ahmed. Analytical stability indicating UPLC assay and validation of Fluticasone propionate in nasal spray inhaler dosage form. World Journal of Pharmaceutical and Life Sciences, 2017; 3, 5: 168-172.
- 25. Anas Rasheed*, Osman Ahmed. Stability indicating UPLC method optimisation and validation of Acetylcysteine in syrup dosage form. European Journal of Pharmaceutical and Medical Research, 2017; 4(7): 485-491.
- 26. Anas Rasheed*, Osman Ahmed. Analytical stability indicating UPLC assay and validation of Ciclesonide in dry powder inhaler dosage form. European Journal of Pharmaceutical and Medical Research, 2017; 4(7): 523-529.
- 27. Anas Rasheed*, Osman Ahmed. Analytical stability indicating UPLC assay and validation of Dextromethorphan in syrup dosage form. European Journal of Pharmaceutical and Medical Research, 2017; 4(7): 548-554.
- 28. Anas Rasheed*, Osman Ahmed. Analytical Development and Validation of a StabilityIndicating Method for the Estimation of Impurities in Budesonide Respules Formulation, International Journal of Applied Pharmaceutical Sciences and Research, 2017; 2(3): 46-54. http://dx.doi.org/10.21477/ijapsr.v2i3.8100
- 29. Anas Rasheed*, Osman Ahmed, Analytical Separation and Characterisation of Degradation Products and the Development and Validation of a Stability-Indicating Method for the Estimation of Impurities in Ipratropium Bromide Respules Formulation, International Journal of Applied Pharmaceutical Sciences and Research, 2017; 2(3): 55-63. http://dx.doi.org/10.21477/ijapsr.v2i3.8101
- Ma, W., Wu, H., & Zheng, H. Analysis of tyrosine kinase inhibitor effects on renal cancer cell metabolism using LC-MS. *Journal of Chromatography B*, 2022; 1208: 123438.
- 31. Mei, Z., Huang, J., & Chen, Z. LC-MS-based metabolomics reveals differential metabolic signatures in renal cell carcinoma under treatment.

- Journal of Proteomics Research, 2021; 20(7): 3215-3226.
- 32. Peng, X., Liu, Y., & Deng, Y. Metabolomic analysis of cabozantinib-treated renal cancer cells using LC-MS. *Cancer Medicine*, 2020; 9(8): 2771-2780.
- 33. Qian, Y., Wang, W., & Zhang, X. Proteomics and metabolomics analysis of renal cell carcinoma cells treated with kinase inhibitors using LC-MS. *Journal of Proteomics*, 2021; 233: 104044.
- Shi, H., Liu, C., & Xu, M. Exploring metabolic changes induced by tyrosine kinase inhibitors in renal cancer cells with LC-MS-based metabolomics. *Journal of Cancer Research*, 2019; 145(3): 523-534.
- 35. Sun, X., Li, H., & Yang, X. Targeted metabolomics of Transitional cell carcinoma using LC-MS reveals potential biomarkers for early detection and treatment monitoring. *Metabolomics*, 2022; 18(5): 35-48.
- Tan, J., Wang, C., & Zheng, L. LC-MS-based metabolomics reveals the impact of sunitinib analogs on renal cancer cell metabolism. *Journal of Chromatography A*, 2020; 1612: 460645.
- 37. Wang, H., Li, Y., & Guo, X. (2021). Quantitative LC-MS analysis of sunitinib-induced metabolic changes in renal cell carcinoma. *Journal of Cancer Metabolism*, 9(2), 134-145.
- 38. Yang, F., & Yu, G. Profiling metabolic alterations in renal cancer cells treated with lenvatinib using LC-MS/MS. *Biochimica et Biophysica Acta (BBA) Molecular Basis of Disease*, 2019; 1865(10): 2636-2645.
- Zhang, L., Chen, S., & Wang, W. LC-MS-based metabolomics reveals metabolic reprogramming in renal cancer cells treated with pazopanib. *Cancer Metabolomics Research*, 2020; 12(6): 256-270.

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