

CADMIUM ACCUMULATION AND ITS EFFECTS ON GROWTH AND CERTAIN METABOLIC ACTIVITIES IN TOMATO (*LYCOPERSICON ESCULENTUM*).

Dr. Shobha Shrivastava*

Department of Botany, S.N.Govt.G.P.G.(Auto)College, Bhopal.

*Corresponding Author: Dr. Shobha Shrivastava

Department of Botany, S.N.Govt.G.P.G.(Auto)College, Bhopal.

Article Received on 28/10/2018

Article Revised on 18/11/2018

Article Accepted on 08/12/2018

ABSTRACT

This paper describes cadmium accumulation and its effects at graded concentration on growth and certain metabolic activities in tomato (*Lycopersicon esculentum*). High soil-Cd levels induced interveinal chlorosis in young expanding leaves which turned to necrosis at later stage of growth. Plant height, total area, total leaf biomass, total plant biomass and fruit yield decreased (up to 74%) in plant grown in Cd added solids. Cadmium supply increased the number of leaves indicating that the tomato plants in response to Cd allocated a greater proportion of photo assimilate for growth and development of photosynthetic organs (leaves). Catalase activity and concentration of chlorophyll, ascorbic acid, N and P declined significantly at higher Cd supply. Cadmium accumulation was greater in root, followed in decreasing order by leaf fruit and shoot. It is concluded that cadmium, in concentrations generally in the contaminated agricultural solid, soils, is inhibitory to growth and metabolism of tomato plants.

KEYWORDS: Cadmium, Photosynthetic organ, Metabolism, Catalase.

INTRODUCTION

Cadmium a common environmental contaminant has various routes of entry into biotic component of the ecosystems. Among other major sources, some agricultural practices such as phosphatic fertilizers, town-refuse composts application, sewage, sludge disposal and mining, smelting, metal refining waste incineration and automobiles add cadmium into the environment. Agricultural soils are mainly contaminated by phosphatic fertilizers, sludge disposal and atmospheric fall out.^[1,2] The accumulations of Cd in the agricultural soils and increased uptake by plants have world-wide concern since occurrence of Cd in the food chain is the most important source for human contamination.^[3] Cadmium is regarded as one of the most toxic metals, although there is no rigid order of toxicity of trace metals in the environment. Being a non-essential element for both plants and animals, there are no critical concentrations below which deficiency of the element would occur. Upper critical concentrations mark the beginning of phytotoxicity. Uptake and translocation of Cd in different plant parts are related to its concentration in the soil, absorption characteristics of species and cultivars, nature of edible parts age of plants and some environmental factors.^[4] The present investigation aimed at studying the accumulation and effects of cadmium on growth and metabolism of tomato

since this is one of the most widely used vegetable crop of the world.

MATERIAL AND METHODS

The present study was designed to examine cadmium accumulation and its effect on tomato plants grown at solid Cd levels generally found in contaminated agricultural solid.^[2] For this purpose, well manured garden solid (pH=7.3 organic carbon=0.88%, available P=0.005% exchangeable K=0.10% cation exchange capacity=16.5 meg/100g) was used. Soil was divided into four classes. Soils of class 1, 2 and 3 were supplemented with 5.00(T₁), 10.0(T₂) and 20.0(T₃) Meg Cd/Kg.respectively, soil of class 4 with no Cd supplement served as control. These soils were filled in the earthen pots of 30 cm diameter (five pots per soil class). Earlier trials with this species indicated that the container size used in these experiments did not restrict root growth. Uniformly sized, twenty day old seedling tomato solid form around the root. Five individuals were harvested immediately to determine an initial fresh to dry weight relationship. After initial observations on height, number of leaves etc. seedlings were planted in earthen pots (1per pot) filled with soils as described above. Samples were collected from the control and each set of treatment on day 45(at 65 days of plant age) and one day 75 (at 95 days of plant age). Fifty percent of plants were harvested at each sampling date. Plant height and number

of leaves, flowers, and fruits were recorded. Chlorophyll pigments were extracted in 80% acetone. The optical densities of extracts were measured at 645 and 663 nm and concentrations determined using the formula given by Mactachtan and Zalik.^[6] For ascorbic acid determination, the method given by Keller and Schwager^[7] was followed. The catalase activity was determined following the method described in Sharma et.al.^[8] For biomass determination. Plants were separated into leaf, stem and root and oven dried separately at 80°C to constant weight. Dry powdered samples were used for the determination of total nitrogen^[9] and phosphorus.^[10] The concentrations of Cd in different plants parts were obtained in oven-dried plant material by atomic

absorption spectrometry after digestion.^[11] The test was performed to test the level of significance difference.

RESULT AND DISCUSSION

Intervinal chlorosis was observed in young leaves of T₂ and T₃ plants at 30 days of growth in Cd supplemented soils. Chlorosis intensified and turned to necrosis in other 30 days. No visible injury symptoms were observed in T₁ plants foliar chlorosis in the recently expanded leaves of tomato due to Cd supply resembled earlier findings.^[12] Plant height decreased significantly at higher Cd levels (**Table: 1**).

Table 1: Effects of varying cadmium levels on development of tomato plants (mean values per plant).

Measurement	Cd (mg/Kg) 65 days				Cd (mg/Kg) 95 days			
	0.00	5.00	10.00	20.00	0.00	5.00	10.00	20.00
Height	52	54	50	4	110	106	82	62
Leaf (No.)	30	32	36	40	31	33	40	44
Flower (No)	-	-	-	-	58	52	35	24
Shoot (gDW)	5.8	5.7	4.1	3.0	13.2	12.5	7.9	6.2
Root (gDW)	1.6	1.9	2.1	1.1	2.2	2.9	1.6	1.1
Fruit (No)	-	-	-	-	-	27	26	148
Total leaf area(cm ²)	815	812	725	605	840	826	706	580
Biomass	10.9	11.1	9.0	6.3	18.7	18.7	12.1	9.3

DW: dry weight

Level of significance: *P<0.05; **P<0.01; ***P<0.001.

Although the number of leaves increased successively in treated plants total leaf area and its biomass decreased significantly in T₂ and T₃ plants. There appeared an initial increase in root biomass successive decline that

finally terminated to significant reduction in total plant biomass. Yield components were significantly reduced in T₂ and T₃ plants (**Table 2**).

Table 2: Effects of varying cadmium levels on chlorophyll, ascorbic acid, catalase activity and Total N and P concentrations in tomato plants.

Measurement	Cd (mg/Kg) 65 days				Cd (mg/Kg) 95 days			
	0.00	5.00	10.00	20.00	0.00	5.00	10.00	20.0
Chlorophyll a	4.60	4.50	3.20	2.60***	3.10	3.10	2.75*	2.51***
Chlorophyll b	3.10	3.05	2.12*	1.62***	2.60	2.65	2.00*	1.38***
Ascorbic acid	11.25	11.90	8.60	6.65***	11.00	11.80	7.50	6.10***
Catalase	4.25	4.40	2.01***	1.60***	4.00	3.95	0.92	0.50
Nitrogen								
Root	10.20	12.00	7.45**	5.85***	9.80	10.00	6.12**	4.10***
Shoot	12.25	2.25	11.80	10.50*	10.30	10.45	9.40	8.70*
Leaf	28.50	28.00	21.20*	15.10***	22.20	20.51	12.60	8.52***
Phosphorus								
Root	3.20	3.51	3.86	2.95	3.00	3.40	2.55*	1.92***
Shoot	4.50	4.45	4.10	3.45**	4.10	4.00	3.44**	2.80**

Values in mg g⁻¹ dry wt. unless mentioned otherwise 1m moles H₂O₂ decomposed /100 mg fresh wt. Level of significance: *P<0.05; **P<0.01; ***P<0.001.

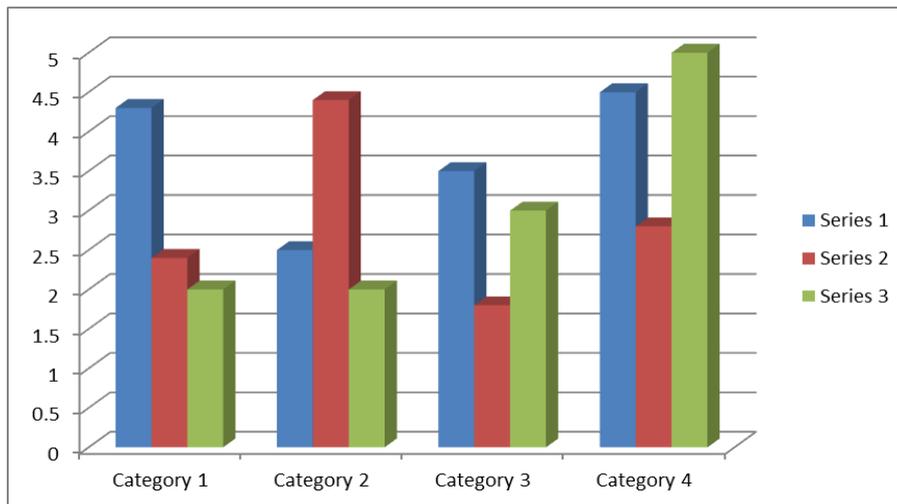


Fig. 1: Effects of varying cadmium levels on chlorophyll, ascorbic acid, catalase activity and Total N and P concentrations in tomato plants.

Ascorbic acid, however, showed a slight increase in T₁ plants. Catalase activity reduced significantly in T₂ and T₃ plant. Total N and P concentrations, except an initial increase in root of T₁ plant decreased in all the treatment compared with the control and the maximum decline was observed in leaf. The decrease in chlorophyll concentration as also reported by Hunter and Vegnano^[14] had a clear impact on plants photosynthetic efficiency and hence on plant biomass accumulation. Ascorbic acid and catalase activity showed a slight improvement at 5 mg/Kg of Cd supply. Poscherieder et.al.^[15] Have

reported increased catalase activity at high Cd supply coincides with the findings in barley^[13] Tomato roots showed an initial increase in biomass Roots of T₁ plants retained N and P at initial stage indicating reduced translocation of these nutrients to above ground plant parts at low level of cadmium.

The tissue concentration of cadmium was greatest in root followed in decreasing order by leaf, fruit and shoot in all treatments. (Table 3.).

Table 3: Cadmium accumulation (µg/g/dry wt.) in different parts of tomato plant grown in soils supplemented with varying of cadmium.

Measurement	Cd(mg/Kg) 65 days				Cd (mg/Kg) 95 days			
	0.00	5.00	10.00	20.00	0.00	5.00	10.00	20.00
Root	ND	9.22	23.54	36.10	ND	13.25	32.65	69.00
		±0.85	±2.10	±2.80		±1.12	±2.80	±5.20
Shoot	ND	3.10	6.95	10.12	ND	5.10	9.20	13.15
		±0.22	±0.50	±1.10		±1.40	±1.75	±1.10
Leaf	ND	7.50	16.20	22.75	ND	11.35	26.20	48.50
		±0.60	±1.12	±2.15		±1.10	±1.80	±3.25
Fruit		ND	-	ND		10.45	26.15	46.18
						±0.92	±1.12	±2.80

ND: Not detectable.

Values are mean ISE for three replicate samples.

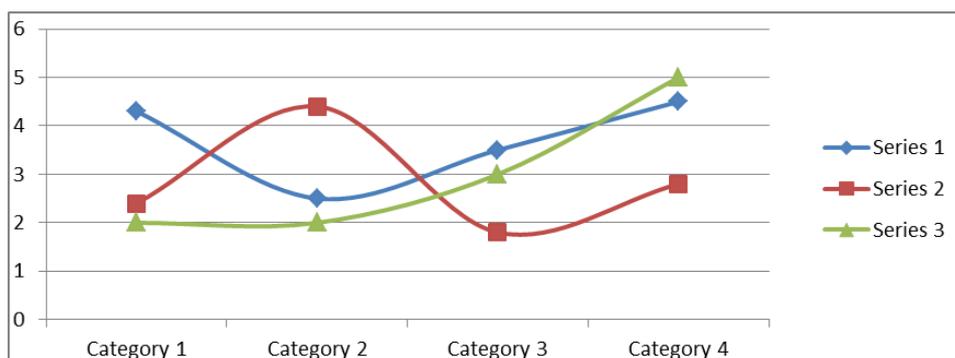


Fig. 2: Cadmium accumulation (µg/g/dry wt.) in different parts of tomato plant grown in soils supplemented with varying of cadmium.

The accumulation of cadmium in different parts of the tomato plant was similar with an earlier report John^[16] reported concentration of Cd in the leaf of eight food crops in soil supplement with 40 mg Cd/Kg ranged from 18.5 mg/Kg (cauliflower) to 264.7 mg/Kg (radish). Jarvis et.al^[17] using 23 different species of crops, observed the Cd accumulation in shoot was significantly low. The study indicated that cadmium in concentrations generally found in agricultural soils receiving phosphates fertilizers and sewage sludge disposal, is inhibitory to growth and metabolism of tomato plants. Furthermore, cadmium is an important constituent of a number of a number of biocides used in agricultural to increase yield. Since tomatoes are used in the cooked vegetables as well as in salads and chatni preparations of Indian recipes, accumulation of cadmium in the fruits may have severe consequences in long-run.

REFERENCES

1. Kurek.E. Czaban J.J.M. Boiling. Sorption of cadmium by micro-organisms in competition with other soil constituents. *App. Environ. Microbiol.*, 1982; 43: 1011-1015.
2. Forstner. U., Cadmium in: Hutzinger.O.(Ed). *The Handbook of Environmental Chemistry.* Springer-Verlag Berlin. Heidelberg, 1980; 3(Part A): 59-107.
3. Bruwanence R.V. Kirchmann and Impens. Cadmium contamination in agriculture and zootechnology *Experimentia*, 1984; 40: 43-52.
4. Pandey J. and Agarwal. Growth responses of tomato to low concentrations of sulphur dioxide and nitrogen dioxide. *Sci.Hort.*, 1994; 58: 67-7.
5. Sharma D.C., Chatterjee C. and Sharma C.P. Chromium accumulation and its effects on wheat (*Triticum aestivum* L.ev. HD 2204) metabolism *Plant Sci.*, 1995; III: 145.
6. Jackson M.L., *Soil Chemical Analysis.* Asia Publishing House. Bombay, 1958; 498.
7. Piper C.S., *Soil and Plant Analysis.* Inter. Sci. Publ.Inc. New York, 1966.
8. Rauser W.E., Early effects of phytotoxic burdens of cadmium, cobalt and nickel and zinc in white beans. *Can. J. Bot.*, 1978; 56: 1744-1749.
9. Agarwal S.C., Bisht S.S. and Sharma C.P., Relative effectiveness of certain heavy metals in producing toxicity and symptoms of iron deficiency in barley. *Can.J.Bot.*, 1977; 55: 1299-1307.
10. Hunter J.G. and Vergnano O., Trace element toxicities in oat plant *Ann. Appl. Biol.*, 1953; 40: 761-777.
11. Poschenieder C. Vazquez M.D. Bonet and Barcelo A., Chromium iron interaction in iron sufficient and iron deficient bean plants. II.Ultrastructural aspects. *J.Plant Nutr.*, 1991; 14: 415-428.
12. John M.K., Cadmium uptake by eight food crops as influenced by various soil levels of cadmium *Environ Pollut.*, 1973; 4: 7-15.
13. Jarvis S.C., Jones L.H. and Hopper M.J., Cadmium uptake from solution and its transport from roots to shoot. *Plant Soil*, 1976; 44: 179-191.
14. T.A. Kipichtechikova, A. Manceau, L. Spadini, F. Panfili, M.A.Marcus and T.Jacquet, "Speciation and solubility of heavy metals in contaminated soil using X-ray microfluorescence, EXAFS spectroscopy, chemical extraction and thermodynamic modeling". *Geochemica ET Cosmochimica Acta*, 2006; 70(9): 2163-2190.
15. D.C. Adriano, *Trace Elements in Terrestrial Environments: Biogeochemistry, Bioavailability and Risks of Metals*, Springer, New York, NY, USA, 2nd edition, 2003.
16. M.J.McLaughlin, B.A.Zarcinas, D.P. Stevens and N.Cook, *Soil testing for heavy metals, Communications in Soil Science and Plant Analysis*, 2000; 31: 11-14, 1661-1700.
17. M.J.McLaughlin, R.E. Hamon, R.G.McLaren, T.W.Speir and S.L.Rogers, "Review: a bioavailability-based rationale for controlling metal and metalloid contamination of agricultural land in Australia and New Zealand", *Australian Journal of Soil Research*, 38: 6.